Amendments to the Specification:

Please replace the paragraph beginning at page 1, line 5 with the following amended paragraph:

This is a non-provisional application which claims priority under 35 U.S.C. § 119(e) to provisional application number 60/228,914 filed on August 29, 2000, and under 35 U.S.C. § 120 of copending U.S. Application Serial No. 09/759,056, filed on January ~~21~~11, 2001, which in turn claims priority to provisional application Nos. 60/175,849 filed on January 13, 2000, 60/197,089 filed on April 14, 2000, and 60/228,914 filed on August 29, 2000, the disclosures of which are hereby expressly incorporated by reference.

Please replace the paragraph beginning at page 19, line 36 with the following amended paragraph:

A "growth inhibitory agent" when used herein refers to a compound or composition which inhibits growth of a cell, especially cancer cell overexpressing any of the genes identified herein, either *in vitro* or *in vivo*. Thus, the growth inhibitory agent is one which significantly reduces the percentage of cells overexpressing such genes in S phase. Examples of growth inhibitory agents include agents that block cell cycle progression (at a place other than S phase), such as agents that induce G1 arrest and M-phase arrest. Classical M-phase blockers include the vincas (vincristine and vinblastine, ~~taxol~~Taxol™, and topo II inhibitors such as doxofubicin, epirubucin, daunorubicin, etoposide, and bleomycin. Those agents that arrest G1 also spill over into S-phase arrest, for example, DNA alkylating agents such as tamoxifen, prednisone, dacarbazine, mechlorethamine, cisplatin, methotrexate, 5-fluorouracil, and ara-C. Further information can be found in The Molecular Basis of Cancer, Mendelsohn and Israel, eds., Chapter 1, entitled "Cell cycle regulation, oncogens, and antineoplastic drugs" by Murakami *et al.*, (WB Saunders: Philadelphia, 1995), especially p. 13.

Please replace the paragraph beginning at page 24, line 24 with the following amended paragraph:

Methods for mRNA extraction are well known in the art and are disclosed in standard textbooks of molecular biology, including Ausubel *et al.*, Current Protocols of Molecular Biology, John Wiley and Sons (1997). Methods for RNA extraction from paraffin embedded tissues are disclosed, for example, in Rupp and Locker, *Lab Invest.* 56:A67 (1987), and De Andrés *et al.*, *BioTechniques* 18:42044 (1995). In particular, RNA isolation can be performed using purification kit, buffer set and protease from commercial manufacturers, such as Qiagen, according to the manufacturer's instructions. For example, total RNA from cells in culture can be isolated using ~~Qiagen~~ Qiagen™ RNeasy min-columns. Total RNA from tissue samples can be isolated using ~~RNA Stat-60~~RNA Stat-60™ (Tel-Test). RNA prepared from tumor can be isolated, for example, by cesium chloride density gradient centrifugation.

Please replace the paragraph beginning at page 25, line 13 with the following amended paragraph:

~~TaqMan~~TAQMAN® RT-PCR can be performed using commercially available equipments, such as, for example, ABI PRIZM 7700™ Sequence Detection System™ (Perkin-Elmer-Applied Biosystems, Foster City, CA, USA), or ~~Lightcycler~~LIGHTCYCLER® (Roche Molecular Biochemicals, Mannheim, Germany). In a preferred embodiment, the 5' nuclease procedure is run on a real-time quantitative PCR device such as the ABI PRIZM 7700™ Sequence Detection System™. The system consists of a thermocycler, laser, charge-coupled device (CCD), camera and computer. The system amplifies samples in a 96-well format on a thermocycler. During amplification, laser-induced fluorescent signal is collected in real-time through fiber optics cables for all 96 wells, and detected at the CCD. The system includes software for running the instrument and for analyzing the data.